



Autocrine regulation of Leydig cell differentiated functions by insulin-like growth factor I and transforming growth factor beta[☆]

Christine Le Roy, Hervé Lejeune, Franck Chuzel, José M. Saez*, Dominique Langlois

INSERM-INRA U 418 and IFREL, Hôpital Debrousse, 69322, Lyon, Cedex 05, France

Abstract

The expression and the maintenance of specific differentiated function of Leydig cells are regulated not only by gonadotropin but by locally produced factors, which may act as autocrine regulators. Many factors, in particular growth factors, have been postulated to have such a type of effect on testicular cells, but very few fulfilled the three criteria required to establish a paracrine/autocrine role: (a) presence of receptors and biological action on local cells; (b) local secretion regulated by physiological signals; and (c) blockade of the factor or its receptors must modify the function of local cells. In the present work we demonstrate that two factors, insulin-like growth factor I (IGF-I) and transforming growth factor β 1 (TGF β 1) fulfilled the three criteria: (a) IGF-I stimulates the transcription of the genes encoding Leydig cell differentiated function, leading to an enhanced steroidogenic responsiveness to LH/hCG; (b) Leydig cells (LC) express and secrete IGF-I and this secretion is enhanced by hCG; and (c) incubation of LC with IgG anti-IGF-I, but not with IgG-control, markedly reduced the steroidogenic responsiveness to LH/hCG. In contrast to IGF-I, TGF β is a potent inhibitor of LC differentiated function. Moreover, LC express TGF β 1 mRNA and secrete this peptide. To prove that the locally produced TGF β has an autocrine role, LC were transfected with 10 μ M of an antisense oligonucleotide (AON) complementary to the translation initiation region of TGF β 1 mRNA. Transfection with AON but not with sense deoxynucleotide induces a complete disappearance of TGF β immunoreactivity in LC and an enhanced hCG-induced testosterone production by LC. This increased steroidogenic responsiveness was associated with a significant enhancement of both LH/hCG receptor mRNA and P450c17 mRNA. Taken together, the above results show that both factors play an autocrine role, although opposite, on Leydig cell function. © 1999 Elsevier Science Ltd. All rights reserved.

1. Introduction

The two main testicular functions are androgen production and spermatogenesis which take place in two different compartments. Steroidogenesis occurs in the vascularized interstitial or intertubular compartments composed of Leydig cells, macrophages, blood vessels and lymphatics and the sheath of myoid or peritubular cells that surrounds the seminiferous tubules. Spermatogenesis occurs within the avascular seminifer-

ous tubule, which is made up exclusively of Sertoli cells and germ cells. The two compartments are separated by the blood–testis barrier which consists of three filters: the surrounding layer of myoid cells, the basement membrane of the tubule and the last and more restricted filter, the Sertoli–Sertoli cell tight junctions, which are established at the time of puberty. This barrier restricts the passage of many factors into the abluminal compartment and lumen of the seminiferous tubule. Thus, all the nutrients, hormones and growth factors required for the very energy-demanding process of spermatogenesis, must be supplied by the ultrafiltrate of interstitial fluid, containing some of the secretory products of Sertoli cells.

Normal function of the testis has long been recognized to be dependent on the pituitary-synthesized gonadotropins. Notwithstanding these requirements

* Proceedings of the Xth International Congress on Hormonal Steroids, Quebec City, Quebec, Canada, 17–21 June 1998.

* Corresponding author. Tel.: +33-4-78-25-18-08; fax: +33-4-78-25-61-68.

E-mail address: saez@lyon151.inserm.fr (J.M. Saez)

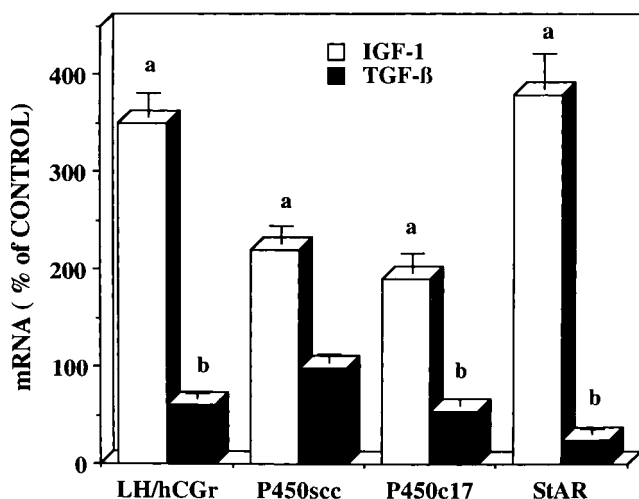


Fig. 1. Effects of IGF-I and TGF β on LH/hCG receptor, P450scc, P450c17 and 3 β -HSD mRNA in porcine Leydig cells. Cells were cultured without or with IGF-I (50 ng/ml) or TGF β (2 ng/ml) for 2 days, and the mRNA levels determined by Northern blot. A letter on a bar indicates a significant difference from the control.

for gonadotropins, numerous reports over the past several years have clearly indicated that locally produced factors may play an important role in the regulation of Leydig cell functions. Recent articles have extensively reviewed this aspect [1–6]. Thus, many factors have been shown to be produced locally and/or to be able to have pleiotropic effects on Leydig cells. This has led to the hypothesis that these factors may play a paracrine/autocrine role on Leydig cell differentiation and function. However, to establish that any factor has such a role, three criteria must be fulfilled: (1) presence of receptors and biological action on local cells; (2) local secretion regulated by physiological signal; and (3) blockade of the factor or its receptor by antibody, antagonist or antisense oligodeoxynucleotides, must modify the function of local cells.

This paper will concentrate on two growth factors, insulin-like growth factor I (IGF-I) and transforming growth factor β 1 (TGF β 1) and demonstrate that in pig Leydig cells these two factors fulfil the three criteria described above and, therefore, that this local production could play an autocrine role in the differentiation and function of Leydig cells.

2. Leydig cells are the site of receptors and action of IGF-I

IGF type I receptor has been demonstrated in Leydig cells of several species by binding studies, cross-linking and immunohistochemistry [7–9]. Moreover, in both pig [8] and rat [9,10] Leydig cells, LH/hCG up-regulates these receptors. In the rat and before puberty, IGF-I stimulates the proliferation of

precursor and immature Leydig cells, and the differentiation of immature into mature Leydig cells [11,12]. Although the steroidogenic action of IGF-I alone, if any, is very small, pretreatment with IGF-I stimulates hCG-supported cAMP and testosterone production by cultured Leydig cells of murine [13,14] and porcine [8,15] origin. The response to cAMP analogs was also enhanced, suggesting that IGF-I may potentiate LH/hCG action at sites both proximal and distal to cAMP generation. Further studies have demonstrated that IGF-I increased LH/hCG receptor number and mRNA, the activity and the mRNA levels of several steroidogenic enzymes [8,16] and StAR (Steroidogenic Acute Regulatory protein) (Fig. 1). The stimulatory effect on these mRNA was mainly at the transcriptional level [16].

Further evidence of the positive role of IGF-I on Leydig cell differentiation and function was obtained by *in vivo* studies. First, in human, isolated GH deficiency [17] or GH resistance, as in the case of Laron syndrome [18], are associated with micropenis, suggesting a decreased fetal Leydig function during the second half of pregnancy, delayed puberty and poor response to exogenous hCG [17] which, in the case of GH deficiency, is very often improved following treatment with GH [17,19]. Second, administration of GH but also of IGF-I to Snell dwarf mice during 7 days increases the number of testicular LH/hCG receptor and the steroidogenic response to hCG [20]. Third, the strongest evidence that IGF-I is crucial in the development and function of Leydig cells came from studies of IGF-I gene knockout mice [21]. In these animals, the testes were reduced in size more than expected from the degree of dwarfism, the number and the volume of Leydig cells were markedly reduced, as well as plasma testosterone levels and the *in vitro* basal and LH-stimulated testosterone production by testicular slices were impaired.

3. Leydig cells are the site of IGF-I production

The expression of IGF-I mRNA has been demonstrated in Leydig cells of several species [21–26]. The levels of IGF-I mRNA in Leydig cells were reduced in hypophysectomized rats but they increased following GH administration [22,24]. These results, however, are contradictory with the lack of effect of human GH on dwarf mouse testicular IGF-I mRNA and on dwarf rat testicular IGF-I peptide [27,28]. Similarly, contradictory results have been reported concerning the role of LH/hCG in testicular IGF-I. *In vivo*, administration of hCG increases IGF-I mRNA levels on rat Leydig cells [23,24], whereas *in vitro*, the opposite effects have been reported [22]. *In vitro*, IGF-I peptide is secreted by rat and porcine Sertoli cells [29,30] as well as by

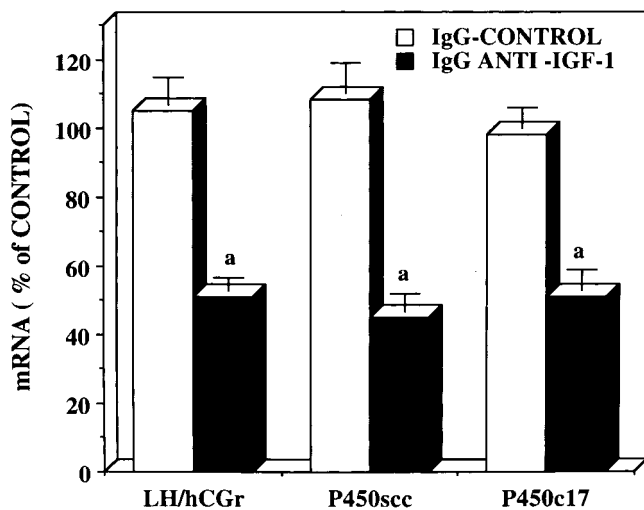


Fig. 2. Effects of IGF-I antiserum on porcine Leydig cells. Cells were treated with 50 μ g of IgG prepared from non-immune serum or IGF-I antiserum for 2 days. At the end of this period, the mRNA levels of LH/hCG receptor, P450scc and P450c17 were evaluated by Northern blot. The results are percent of cells cultured in the same medium without IgG. A letter indicates a significant difference from the control.

Leydig cells [31,32] of the same species, and this secretion was stimulated in a dose-dependent fashion by hCG and FSH, respectively [32].

4. Inhibition of secreted IGF-I by antibodies inhibits Leydig cell functions

Although endogenous secreted IGF-I plays a positive trophic effect on Leydig cell functions, inhibition of its action by specific antibodies should produce the opposite effects. Treatment of rat Leydig cells with IGF-I antiserum decreased the steroidogenic responsiveness to LH, when compared to cells treated with non-immune serum [33]. Moreover, the IGF-I antiserum almost completely blocked the positive action of Sertoli cells on Leydig cell steroidogenic capacity. Similarly, treatment of porcine Leydig cells with increasing concentrations of IgG prepared from IGF-I antiserum decreased the steroidogenic responsiveness to hCG stimulation, whereas the response of cells treated with the same amounts of IgG prepared from non-immune serum had the opposite effect [34]. This decrease in the steroidogenic responsiveness of Leydig cells treated with IgG anti-IGF-I was associated with a decrease of LH/hCG receptor, P450scc and P450c17 mRNA levels (Fig. 2). Taking together the *in vivo* and *in vitro* results, it appears that IGF-I is one of the factors for which there is convincing evidence to postulate that, in addition to its endocrine role, this factor plays a paracrine/autocrine role in the regulation of Leydig cells.

5. Leydig cells are the site of reception and action of TGF β

Porcine Leydig cells contain specific TGF β receptors of high affinity [35] and the developmental testicular expression of mRNAs for the three types of receptor has been analyzed in the rat [36]. TGF β has pleiotropic effects on Leydig cells. Treatment with TGF β reduced hCG-induced testosterone production in pig [37,38], rat [39,40] and mouse [41] Leydig cells. In both pig [37] and rat [39] Leydig cells, TGF β reduced also the steroidogenic response to cAMP derivatives, suggesting an impairment of the steroidogenic pathway. Further studies have shown that in pig Leydig cells TGF β causes a decrease of LH/hCG receptor number and mRNA as well as of P450c17 and StAR mRNA [16] (see also Fig. 1).

6. Leydig cells are the site of production of TGF β

Several groups have demonstrated that rat, mouse and pig LC express TGF β s mRNAs [42–45] and TGF β -like immunoreactive material [45–49]. In the rat [43] and the mouse testis [44], TGF β 1 and TGF β 3 have been shown to be expressed in Sertoli cells and peritubular cells throughout testicular development. In immature pig testis, both Leydig cells and Sertoli cells express TGF β 1 mRNA and protein [45]. During fetal life, immunostaining for TGF β 1 appears in the rat primordial Sertoli cells on fetal day 14.5, increases until day 16.5, and becomes faint from fetal day 18.5 onward, whereas in fetal Leydig cells a positive reaction for TGF β 1 appeared on day 16.5 and became very intense during late fetal life and persisted until postnatal day 20 [48]. Contradictory results have been reported concerning the presence of immunoreactivity for TGF β 1 in adult rat Leydig cells, disappearance at the time of puberty [47], or positive but slight staining [48]. This discrepancy may result from differences in the antibodies used.

In vitro studies have shown that rat Sertoli cells and peritubular cells secrete the three TGF β isoforms [43], whereas pig Sertoli cells and Leydig cells secrete mainly TGF β 1 [45]. In cultured Sertoli cells from 20-day-old rats, FSH reduces TGF β 2 mRNA and protein [43], whereas in pig Sertoli cells FSH decreases TGF β 1 mRNA and protein [45]. In contrast, in rat fetal testis, FSH stimulates TGF β 1 protein secretion, but not its mRNA levels and these effects were potentiated by LH [46].

Taken together, the above results clearly demonstrate that testicular somatic cells express and secrete TGF β s, but there is some diversity in their regulation according to the isoform, the species and/or the devel-

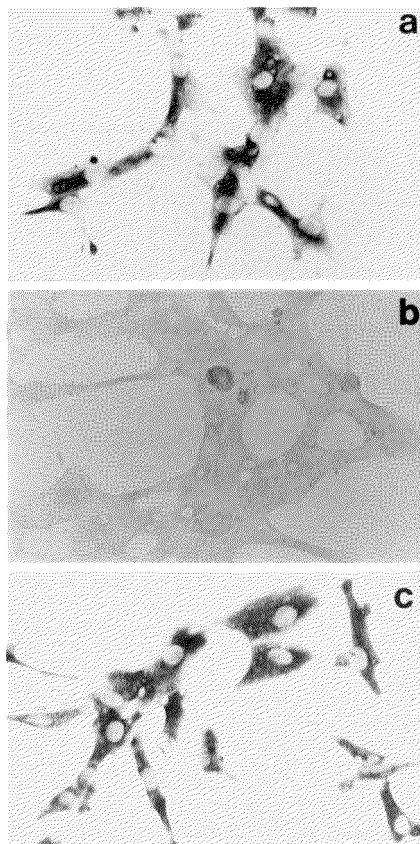


Fig. 3. TGF β 1 protein content in control (a) AON (b) and SON (c) treated Leydig cells. The immunocytochemical staining was performed using a specific TGF β 1 antibody.

opment stage. However, Leydig cells express and secrete predominantly TGF β 1.

7. Inhibition of TGF β 1 synthesis in Leydig cells increases their steroidogenic activity

This study has used two approaches to block the effects of endogenous secreted TGF β 1 in Leydig cells: TGF β 1 antibodies and oligonucleotide antisense. The first approach did not give clear-cut results probably because the TGF β 1 antibody used recognizes only the active form of this peptide [45] but Leydig cells, as many other cells, secrete the peptide in a latent form [45]. For the second approach, this study used antisense (AON) and sense (SON) unmodified 15-bases deoxyribonucleotides, corresponding to the translation initiation region of TGF β 1 mRNA. Cells were transfected for 24 h by a cationic liposome-mediated transfection method, as described before [50] and cultured for 44 h.

Neither AON nor SON modified TGF β mRNA levels (data not shown). In contrast, AON, but not SON, completely blocked TGF β 1 protein synthesis as revealed by immunocytochemistry (Fig. 3).

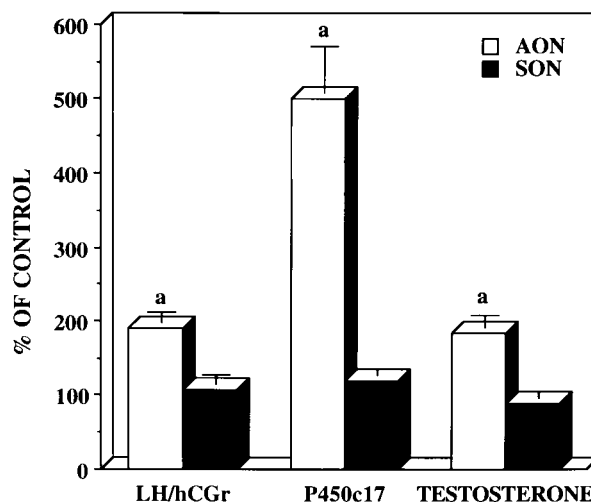


Fig. 4. Effects of antisense (AON) and sense (SON) on LH/hCG receptor mRNA, P450c17 mRNA and hCG-induced testosterone production. Cells were transfected for 24 h without or with 10 μ M of AON or SON. After 44 h of culture, RNA was extracted and analyzed by Northern blot. Other cells were stimulated with hCG (10^{-9} M) and the testosterone production measured after 2 h. A letter indicates a significant difference from control.

To investigate the biological consequences of TGF β 1 protein synthesis inhibition in Leydig cells, two types of parameters were measured. First, this study evaluated the mRNA levels of LH/hCG receptor and P450c17, since, as indicated above (Fig. 1), exogenous TGF β 1 reduced both. The results of Fig. 4 clearly show that TGF β 1 AON, but not SON, increased LH/hCG receptor and P450c17 mRNA levels by about 2- and 5-fold (Fig. 4). Second, the steroidogenic responsiveness to hCG was studied and it was demonstrated that AON, but not SON, increased by about 1.6-fold the hCG-induced testosterone production (Fig. 4).

All these data show that the inhibition of TGF β 1 synthesis by AON produces effects which are opposite to that induced by exogenous TGF β 1 in Leydig cells.

8. Conclusions

Leydig cells from several species contain specific

Table 1
Effects of TGF β and IGF-I on Leydig cells differentiated functions

	TGF β 1	IGF-I
hCG R mRNA	↓	↑
StAR mRNA	↓	↑
P450 _{scc} mRNA	→	↑
P450 17 α mRNA	↓	↑
hCG-induced testosterone production	↓	↑

receptors from both IGF-I and TGF β and these peptides are able to regulate, but in opposite manner (Table 1), the expression of several specific Leydig cell functions. In addition, Leydig cells express and secrete both IGF-I and TGF β and their secretion is regulated by physiological signals. Finally, the inhibition of action or synthesis of these peptides caused the opposite effects to those produced by the addition of these factors. In addition to these autocrine effects, it is likely that these factors also play a paracrine role, since both are also secreted by the other somatic testicular cells. In vivo studies in human and experimental animals have confirmed the crucial role of IGF-I on Leydig cell differentiation and function, but such studies are still required to confirm the physiological relevance of TGF β s in the regulation of Leydig cell function.

Acknowledgements

This work was supported by a grant from Biologie Humaine, Université Claude-Bernard. The authors thank J. Bois and M.A. di Carlo for their secretarial help.

References

- [1] M.K. Skinner, Cell–cell interactions in the testis, *Endocrine Reviews* 12 (1991) 45–77.
- [2] J.F. Ackland, N.B. Schwartz, K.E. Mayo, R.E. Dodson, Nonsteroidal signals originating in the gonads, *Physiological Reviews* 72 (1992) 731–788.
- [3] R.M. Sharpe, Experimental evidence for Sertoli–germ cell and Sertoli–Leydig cell interactions, in: L.D. Russell, M.D. Griswold (Eds.), *The Sertoli Cell*, Cache River Press, Clearwater, FL, 1993, pp. 391–418.
- [4] J.M. Saez, Leydig cells: endocrine, paracrine, and autocrine regulation, *Endocrine Reviews* 15 (1994) 574–626.
- [5] J.M. Saez, H. Lejeune, Regulation of Leydig cell function by hormones and growth factors other than LH and IGF-I, in: A.H. Payne, M.P. Hardy, L.D. Russell (Eds.), *The Leydig Cell*, Cache River Press, Vienna, IL, 1996, pp. 383–406.
- [6] L. Gnassi, A. Fabbri, G. Spera, Gonadal peptides as mediators of development and functional control of the testis: an integrated system with hormones and environment, *Endocrine Reviews* 18 (1997) 541–609.
- [7] B.G. Vanelli, A. Natali, T. Barni, M. Serio, C. Orlando, G. Balboni, Insulin-like growth factor-I (IGF-I) and IGF-I receptor in human testis: an immunohistochemical study, *Fertility and Sterility* 49 (1988) 666–669.
- [8] M.H. Perrard-Sapori, P.G. Chatelain, C. Jaillard, J.M. Saez, Characterization and regulation of somatomedin-C/insulin-like growth factor I (Sm-C/IGF-I) receptors on cultured pig Leydig cells. Effects of Sm-C/IGF-I on luteotropin receptors and steroidogenesis, *European Journal of Biochemistry* 165 (1987) 209–214.
- [9] T. Lin, J. Blaisdell, J.F. Haskell, Hormonal regulation of type I insulin-like growth factor receptors of Leydig cells in hypophysectomized rats, *Endocrinology* 123 (1988) 134–139.
- [10] M.L. Nagpal, D.L. Wang, J.H. Calkins, W.W. Chang, L. Tu, Human chorionic gonadotropin up-regulates insulin-like growth factor-I receptor gene expression of Leydig cells, *Endocrinology* 129 (1991) 2820–2826.
- [11] S. Khan, K. Teerds, J. Dorrington, Growth factor requirements for DNA synthesis by Leydig cells from the immature rat, *Biology of Reproduction* 46 (1992) 335–341.
- [12] A. Moore, I.D. Morris, The involvement of insulin-like growth factor I in local control steroidogenesis and DNA synthesis of Leydig and non-Leydig cells in the rat testicular interstitium, *Journal of Endocrinology* 138 (1993) 107–114.
- [13] T. Lin, J. Haskell, N. Vinson, L. Terracio, Characterization of insulin and insulin-like growth factor I receptors of purified Leydig cells and their role in steroidogenesis in primary culture: a comparative study, *Endocrinology* 119 (1986) 1641–1647.
- [14] S.J. Gelber, M.P. Hardy, S.M.L.C. Mendishandagama, S.J. Casella, Effects of insulin-like growth factor-I on androgen production by highly purified pubertal and adult rat Leydig cells, *Journal of Andrology* 13 (1992) 125–130.
- [15] M. Bernier, P. Chatelain, J.P. Mather, J.M. Saez, Regulation of gonadotropin receptors, gonadotropin responsiveness, and cell multiplication by somatomedin-C and insulin in cultured pig Leydig cells, *Journal of Cellular Physiology* 129 (1986) 257–263.
- [16] F. Chuzel, A.M. Clark, O. Avallet, J.M. Saez, Transcriptional regulation of the lutropin/human choriogonadotropin receptor and three enzymes of steroidogenesis by growth factors in cultured pig Leydig cells, *European Journal of Biochemistry* 239 (1996) 8–16.
- [17] H.E. Kulin, E. Samdjlake, R. Santen, S. Santner, The effects of growth hormone on the Leydig cell response to chorionic gonadotropin in boys with hypopituitarism (1981) 45, 468–472.
- [18] Z. Laron, Laron-type dwarfism (hereditary somatomedin deficiency). A review, *Advances in Internal Medicine and Pediatrics* 51 (1984) 117–140.
- [19] M.A. Rivarola, J.J. Heinrich, E.J. Podesta, M.F. Chondjnik, C. Bergada, Testicular function in hypopituitarism, *Pediatric Research* 6 (1972) 634–641.
- [20] P.G. Chatelain, P. Sanchez, J.M. Saez, Growth hormone and insulin-like growth factor-I treatment increase testicular luteinizing hormone receptors and steroidogenic responsiveness of growth hormone deficient dwarf mice, *Endocrinology* 128 (1991) 1857–1862.
- [21] J. Baker, M.P. Hardy, J. Zhou, C. Bondy, F. Lupu, A.R. Bellve, A. Efstratiadis, Effects of an IGF1 gene null mutation on mouse reproduction, *Molecular Endocrinology* 10 (1996) 903–918.
- [22] T. Lin, D.L. Wang, J.H. Calkins, H. Guo, R. Chi, P.R. Housley, Regulation of insulin-like growth factor-I messenger ribonucleic acid expression in Leydig cells, *Molecular and Cellular Endocrinology* 73 (1990) 147–152.
- [23] A. Moore, C.L.C. Chen, J.R.E. Davis, I.D. Morris, Insulin-like growth factor-I mRNA expression in the interstitial cells of the rat testis, *Journal of Molecular Endocrinology* 11 (1993) 319–324.
- [24] J. Closset, A. Gothot, B. Sente, M.L. Scippo, A. Igout, M. Vandenbroeck, D. Dombrowicz, G. Hennen, Pituitary hormones dependent expression of insulin-like growth factors I and II in the immature hypophysectomized rat testis, *Molecular Endocrinology* 3 (1989) 1125–1131.
- [25] A.M. Clark, S.E. Samaras, J.M. Hammond, D.R. Hagen, Changes in the messenger ribonucleic acid for insulin-like growth factor-I and -II in the porcine testis during and between two waves of testicular development, *Biology of Reproduction* 50 (1994) 993–999.
- [26] H. Lejeune, P. Sanchez, J.M. Saez, Enhancement of long-term testosterone secretion, and steroidogenic enzyme expression in

- human Leydig cells by coculture with human Sertoli cell-enriched preparation. *International Journal of Andrology* (1998) in press.
- [27] L.S. Mathews, G. Norstedt, R.D. Palmiter, Regulation of insulin-like growth factor I gene expression by growth hormone. *Proceedings of the National Academy of Sciences of USA* 83 (1986) 9343–9347.
- [28] J. Spiteri-Grech, J.M.S. Bartlett, E. Nieschlag, Regulation of testicular insulin-like growth factor-I in pubertal growth hormone-deficient male rats, *Journal of Endocrinology* 131 (1991) 279–285.
- [29] E.P. Smith, M.E. Svoboda, J.J. Van Wyk, A.L. Kierszenbaum, L.L. Tres, Partial characterization of a somatomedin-like peptide from the medium of cultured rat Sertoli cells, *Endocrinology* 120 (1987) 186–193.
- [30] P.G. Chatelain, D. Naville, J.M. Saez, Somatomedin-C/insulin-like growth factor I-like material secreted by porcine Sertoli cells in vitro: characterization and regulation, *Biochemical and Biophysical Research Communications* 146 (1987) 1009–1017.
- [31] J. Cailleau, S. Vermeire, G. Verhoeven, Independent control of the production of insulin-like growth factor-I and its binding protein by cultured testicular cells, *Molecular and Cellular Endocrinology* 69 (1990) 79–89.
- [32] D. Naville, P.G. Chatelain, O. Avallet, J.M. Saez, Control of production of insulin-like growth factor-I by pig Leydig and Sertoli cells cultured alone or together. Cell-cell interactions, *Molecular and Cellular Endocrinology* 70 (1990) 217–224.
- [33] G. Verhoeven, J. Cailleau, Influence of coculture with Sertoli cells on steroidogenesis in immature rat Leydig cells, *Molecular and Cellular Endocrinology* 71 (1990) 239–251.
- [34] H. Lejeune, R. Habert, J.M. Saez, Origin, proliferation and function of Leydig cells, *Journal of Molecular Endocrinology* 20 (1998) 1–28.
- [35] J.M. Saez, O. Avallet, D. Naville, M.H. Perrard-Sapori, P.G. Chatelain, Sertoli-Leydig cell communications, *Annals of the New York Academy of Sciences* 564 (1989) 210–231.
- [36] B. LeMagueresse-Battistoni, A.M. Morera, I. Goddard, M. Benahmed, Expression of mRNAs for transforming growth factor-beta receptors in the rat testis, *Endocrinology* 136 (1995) 2788–2791.
- [37] O. Avallet, M. Vigier, M.H. Perrard-Sapori, J.M. Saez, Transforming growth factor beta inhibits Leydig cell functions, *Biochemical and Biophysical Research Communications* 146 (1987) 575–581.
- [38] A.M. Morera, C. Cochet, M. Keramidas, M.A. Chauvin, E. De Peretti, M. Benahmed, Directing regulating effects of transforming growth factor beta on the Leydig cell steroidogenesis in primary culture, *Journal of Steroid Biochemistry* 30 (1988) 443–447.
- [39] T. Lin, J. Blaisdell, J.F. Haskell, Transforming growth factor beta inhibits Leydig cell steroidogenesis in primary culture, *Biochemical and Biophysical Research Communications* 146 (1987) 387–394.
- [40] C. Gautier, C. Levacher, J.M. Saez, R. Habert, Transforming growth factor beta 1 inhibits steroidogenesis in dispersed fetal testicular cells in culture, *Molecular and Cellular Endocrinology* 131 (1997) 21–30.
- [41] N.M. Van Bebakar, J.W. Honour, D. Foster, Y.L. Liu, H.S. Jacobs, Regulation of testicular function by insulin and transforming growth factor β , *Steroids* 55 (1990) 266–270.
- [42] M.K. Skinner, H.L. Moses, Transforming growth factor beta gene expression and action in the seminiferous tubule: peritubular cell-Sertoli cell interactions, *Molecular Endocrinology* 3 (1989) 625–634.
- [43] B.P. Mullaney, M.K. Skinner, Transforming growth factor- β (β 1, β 2, and β 3) gene expression and action during pubertal development of the seminiferous tubule. Potential role at the onset of spermatogenesis, *Molecular Endocrinology* 7 (1993) 67–76.
- [44] T. Watrin, L. Scotto, R.K. Assoian, D.J. Wolgemuth, Cell lineage specificity of expression of the murine transforming growth factor- β 3 and transforming growth factor- β 1 genes, *Cell Growth and Differentiation* 2 (1991) 77–83.
- [45] O. Avallet, M. Vigier, P. Leduque, P.M. Dubois, J.M. Saez, Expression and regulation of transforming growth factor- β 1 messenger ribonucleic acid and protein in cultured porcine Leydig and Sertoli cells, *Endocrinology* 134 (1994) 2079–2087.
- [46] C. Gautier, C. Levacher, J.M. Saez, R. Habert, Expression and regulation of transforming growth factor beta 1 mRNA and protein in rat fetal testis in vitro, *Biochemical and Biophysical Research Communications* 236 (1997) 135–139.
- [47] K.J. Teerds, J.H. Dorrington, Localization of transforming growth factor β 1 and β 2 during testicular development in the rat, *Biology of Reproduction* 48 (1993) 40–45.
- [48] C. Gautier, C. Levacher, O. Avallet, M. Vigier, V. Rouiller-Fabre, L. Lecerf, J. Saez, R. Habert, Immunohistochemical localization of transforming growth factor- β 1 in the fetal and neonatal rat testis, *Molecular and Cellular Endocrinology* 99 (1994) 55–61.
- [49] R. Olaso, C. Gautier, C. Levacher, P. Durand, J. Saez, R. Habert, The immunohistochemical localization of transforming growth factor- β 2 in the fetal and neonatal rat testis, *Molecular and Cellular Endocrinology* 126 (1997) 165–172.
- [50] C. LeRoy, P. Leduque, P.M. Dubois, J.M. Saez, D. Langlois, Repression of transforming growth factor β 1 protein by antisense oligonucleotide-induced increase of adrenal cell differentiated functions, *Journal of Biological Chemistry* 271 (1996) 11,027–11,033.